

# **truChIP™ Ultra-Low Chromatin Shearing Kit**

Adaptive Focused Acoustics™ (AFA)-based chromatin shearing for  
ChIP-based applications

Products PN 520156 and PN 520158



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## INTENDED USE

The truChIP™ Kit is intended for use in research applications (RUO). This product is not intended for the diagnosis, prevention, or treatment of disease.

## INTRODUCTION

The truChIP™ Ultra-Low Chromatin Shearing Kit is optimized for the efficient and reproducible shearing of chromatin from adherent and suspension mammalian cell lines using Covaris AFA™ Focused-ultrasonicators. Focused-ultrasonicators provide a non-contact and isothermal method of shearing chromatin without compromising the structural integrity of the target epitopes of interest. This kit can be used to prepare sample for ChIP-qPCR, ChIP-ChiP, and ChIP-Seq applications.

truChIP™ is compatible for use with a variety of cell lines, however, additional optimization for your specific cell line may improve results. To start, Covaris recommends for users to perform a one-time fixation and shearing time course study to empirically determine the optimal treatment conditions.

## REVISION HISTORY

Part Number	Revision	Date	Description of change
010255	C	04/17	Update template and publish ME220 settings
010255	D	7/17	Remove specific content information for Buffer D3
010255	E	7/17	Correct procedure overview

## PROCEDURE OVERVIEW

**Collect Cells and resuspend in fixing buffer**



**Crosslink DNA-proteins with formaldehyde**



**Add Covaris shearing buffer**



**Shear chromatin with an AFA™ Focused Ultrasonicator**



**Dilute with IP buffer and IP**

## SAMPLE INPUT REQUIREMENTS

The truChIP™ Ultra Low Chromatin Shearing Kit is designed to perform efficient chromatin shearing of 100,000 cells or less.

Single Sample	Low Cell
Input cell number	<100,000
Number of samples sheared per kit	50
AFA tube	microTUBE–130
Shearing volume	130 µl

## KIT CONTENTS

Buffer A	4 mL	10X Fixing Buffer
Buffer D3	6 mL	10X SDS Shearing Buffer
Buffer E	6 mL	1X Quenching Buffer
Buffer F	0.8 mL	100X Protease Inhibitor Cocktail
Formaldehyde	2x1 mL ampules (PN 520156 only)	16% methanol-free formaldehyde

Safety Data Sheets: <http://covarisinc.com/resources/safety-data-sheets>

## STORAGE

The kit is shipped cold and should be stored at 2-8 °C. Prior to use, kit reagent Buffers D3 and E may have to be warmed to 55 °C to dissolve precipitate and then cooled to room temperature before use.

**Note:** Mix solutions well before use to ensure solutions are completely solubilized

## SUPPLIED BY USER

- Molecular Biology Grade Water – Thermo Scientific (Cat. No. SH3053802), Mo Bio (Cat. No. 17012-200), or equivalent
- Phosphate Buffered Salt Solution (PBS) – Mo Bio (Cat. No. 17330-500), Thermo Scientific (Cat. No. SH30256.FS), or equivalent
- RNase A (DNase free) Thermo Scientific (Cat. No. EN0531) or equivalent
- Proteinase K (RNase and DNase free) Thermo Scientific (Cat. No. 17916), NEB (Cat. No. P8102S), or equivalent
- Covaris Focused Ultrasonicator (M220, ME220, S220, E220 Evolution, E220, or LE220)
- Refrigerated centrifuge having 15,000 x g capability
- Rocker – Nutator® or equivalent
- AFA tubes & holders/racks

### Ultra-Low Cell Protocol (<100K) Consumables & Holders/Racks Required

Part Number	Description	M220 Holder & Insert	ME220 Holder & Insert	S-Series Holder	E220Evo Rack	E220 Rack	LE220 Rack
520045	microTUBE AFA Fiber Pre-Slit Snap-Cap	500414 & 500489	500514 & 500526	500114	500433	500111	NA
520052	microTUBE AFA Fiber Crimp-Cap	NA	500514 & 500526	500114	500433	500282	500282
520216	microTUBE-130 AFA Fiber Screw-Cap	500414 & 500489	500522 & 500534	500339	NA	NA	NA
520053	8 microTUBE Strip V1	NA	500514 & 500526	NA	500430	500191	500191
520217	8 microTUBE-130 AFA Fiber Strip V2	NA	500518 & 500526	NA	NA	NA	NA
520078	96 microTUBE Plate	NA	NA	NA	NA	No rack required	500329

# PROTOCOL

## A. Cell Preparation and Crosslinking

Follow the Cell Preparation and Crosslinking method (**A.1 – Suspension** and **A.2 – Adherent**) for your cell culture type.

**Note:** ChIP assays are sensitive to crosslinking and shearing conditions. Therefore, Covaris recommends users to include multiple fixation and shearing time points to empirically determine the optimal treatment conditions.

### A.1 Suspension cells

1. Prepare solutions for the appropriate number of samples being processed fresh before starting.

Buffer	Instructions
<b>1X Cold PBS</b>	<b>Final Volume: 2.0 mL per sample</b> - Store on ice
<b>1X Fixing Buffer A</b>	<b>Final Volume: 0.5 mL per sample</b> - Mix 50 µl of Fixing Buffer A with 0.450 mL of molecular biology grade water
<b>Fresh 5% Formaldehyde</b>	<b>Final Volume: 1 mL per 1 to 20 samples</b> - Mix 312.5 µl of 16% Fresh Formaldehyde with 687.5 µl of molecular biology grade water
<b>Quenching Buffer E</b>	Place in a 55°C water bath to dissolve crystals, then place at ambient

**Note:** The use of fresh methanol-free formaldehyde is required to achieve reproducible results. The methanol-free formaldehyde ampule is for one-time use only—storage for later use is not recommended

2. Collect cells by centrifugation at 200 x g for 5 minutes at room temperature. Remove media and wash cells once with 1X Cold PBS and collect cells again by centrifugation.

Reagent	Ultra-Low Cell
<b>1X Cold PBS</b>	400 µl
<b>Input cell number</b>	1x10 <sup>5</sup> Cells
<b>Centrifuge Tube</b>	1.5 mL

**Note:** Some cells do not pellet well at 200 x g. If a “spongy” pellet is not visible, increase speed at 100 x g intervals until a pellet is visible.

3. Resuspend cells in room temperature 1X Fixing Buffer A.

Reagent	Ultra-Low Cell
Fixing Buffer A	400 $\mu$ l

4. Crosslink cells by adding freshly prepared 5% formaldehyde solution to a final concentration of 0.25% and set timer.

Reagent
Fresh 5% Formaldehyde

**Note:** The use of fresh methanol-free formaldehyde is required to achieve reproducible results.

5. Place cells on a rocker at room temperature for the recommended time.

**Note:** We recommend including two fixation times. Typically, **2.5 and 5 minutes** for stem and primary cells, and **5 and 10 minutes** for all other cell types.

6. Quench the crosslinking reaction by adding the appropriate volume of Quenching Buffer E to the fixed cells. Keep cells on a rocker at room temperature for an additional 5 minutes.

Reagent	Ultra-Low Cell
Quenching Buffer E	12 $\mu$ l

7. Collect cells by centrifuging at 500 x g for 5 minutes at room temperature.

8. Aspirate the supernatant and wash twice with cold PBS.

Reagent	Ultra-Low Cell
1X Cold PBS	300 $\mu$ l

9. Collect cells by centrifugation at 500 x g for 5 minutes, 4 °C.

10. Proceed to nuclei preparation and chromatin shearing steps.

**Note:** You may flash-freeze the fixed cells in liquid nitrogen at this point and store at -80 °C for short periods of time (*e.g.*, 2 to 3 days). Longer-term storage is not recommended.



## A.2 Adherent cells

1. Grow the proper amount of cells to conduct a single ChIP assay or the initial time course until they are 80 to 90% confluent.
2. Prepare solutions for the appropriate number of samples being processed fresh before starting.

Buffer	Instructions
<b>1X cold PBS</b>	<b>Final Volume: 14 mL</b> - Store on ice
<b>1X Fixing Buffer A</b>	<b>Final Volume: 2 mL</b> - Mix 200 µl of Fixing Buffer A with 1.8 mL of molecular biology grade water
<b>Fresh 5% Formaldehyde</b>	<b>Final Volume: 1 mL</b> - Mix 312.5 µl of 16% Fresh Formaldehyde with 687.5 µl of molecular biology grade water
<b>Quenching Buffer E</b>	Place in a 55°C water bath to dissolve crystals, then place at ambient
<b>Important Notes</b>	
<ul style="list-style-type: none"> <li>• The use of fresh methanol-free formaldehyde is required to achieve reproducible results</li> <li>• The methanol-free formaldehyde ampule is for one-time use only—storage for later use is not recommended</li> </ul>	

3. Remove media and wash each plate one time with cold PBS.

Reagent	Volume
<b>1X PBS</b>	2 mL

4. Remove PBS and add room temperature 1X Fixing Buffer A to each dish.

Reagent	Volume
<b>Fixing Buffer A</b>	2 mL

5. Crosslink cells by adding freshly prepared 5% formaldehyde solution to a final concentration of 0.25% and start timing the crosslinking reaction.

Reagent	Volume
<b>Fresh 5% Formaldehyde</b>	100 µl

**Note:** We recommend including two fixation times. Typically, **2.5 and 5 minutes** for stem and primary cells, and **5 and 10 minutes** for all other cell types.

6. Place cells on a rocker at room temperature for the recommended time.

**Note:** The use of fresh methanol-free formaldehyde is required to achieve reproducible results.

7. Quench the crosslinking reaction by adding the appropriate volume of Quenching Buffer E to the fixed cells. Keep cells on a rocker at room temperature for an additional 5 minutes.

Reagent	Ultra-Low Cell
Quenching Buffer E	60 $\mu$ l

8. Completely aspirate the solution from the plate.

9. Add cold PBS to each dish and scrape cells from the plate into the proper vessel.

Reagent	Ultra-Low Cell
1X Cold PBS	450 $\mu$ l
Centrifuge Tube	1.5 mL tube

10. Wash the plate with an additional volume of cold PBS to collect any remaining cells.

Reagent	Ultra-Low Cell
1X Cold PBS	450 $\mu$ l
Centrifuge Tube	1.5 mL tube

11. Collect cells by centrifuging at 500 x g for 5 minutes, 4°C.

**Note:** Some cells do not pellet well at 200 x g. If a “spongy” pellet is not visible, increase speed at 100 x g intervals until a pellet is visible.

12. Wash cells twice by resuspending in cold PBS and collecting by centrifugation at 500 x g, 4°C.

Reagent	Ultra-Low Cell
1X Cold PBS	450 $\mu$ l
Centrifuge Tube	1.5 mL tube

13. Completely and carefully aspirate the supernatant from the tube(s) and place tube(s) on ice.

14. Proceed to chromatin shearing steps.

**Note:** You may flash-freeze the fixed cells in liquid nitrogen at this point and store at -80 °C for short periods of time (*e.g.*, 2 to 3 days). Longer-term storage is not recommended.

## B. Chromatin Shearing

1. Resuspend nuclei pellet in the Shearing Buffer D3 and transfer to appropriate AFA Tube(s). If conducting a shearing time course experiment aliquot 130  $\mu$ l of  $\leq 100,000$  fixed cells into 6 microTUBES

Reagent	Ultra-Low Cell
Shearing Buffer D3	130 $\mu$ l
AFA Tube	microTUBE-130

2. Shear chromatin with an AFA Focused-ultrasonicator with appropriate rack or holder; settings are provided in **Appendix A**. For the shearing time course, use processing times of 2, 4, 6, 8, 10, and 12 minutes.

**Note:** Optimization of shearing time should be conducted whenever experimental parameters (*e.g.*, cell type, cell number, or sample volumes) are changed.

3. After shearing, transfer samples into a pre-chilled microcentrifuge tube and place on ice until all tubes are processed. If batch processing using a high-throughput ultrasonicator (*e.g.* E220), samples can be maintained in the instrument's water bath at 4 °C before and after processing.
4. After processing, add 2-3 volumes of your IP buffer, and centrifuge samples at 10,000 x g, 4°C for 5 minutes to pellet insoluble material
5. Transfer the supernatant to a new pre-chilled microcentrifuge tube.

**Note 1:** To check the efficiency of your shearing, reserve 130  $\mu$ l of the sheared chromatin and see **Appendix B** for detailed instructions

**Note 2:** Sheared chromatin can be stored at 4 °C for up to 2 days

**Note 3:** Freezing sheared chromatin is not recommended. Freeze/thaw cycles reduce IP efficiency and reproducibility

**Note 4:** For subsequent immunoprecipitation, sheared chromatin can be diluted in the desired immunoprecipitation buffer. Alternatively, the composition of the shearing buffer can be adjusted appropriately for immunoprecipitation. **The 1 $\times$  SDS Shearing Buffer D3 composition is: 10 mM Tris-HCl pH 7.6, 1 mM EDTA, 0.1% SDS**

## SUPPLEMENTAL MATERIAL

### Appendix A: AFA Focused-ultrasonicator Operating Conditions

Ultra-Low Cell Chromatin Shearing Protocol						
Instrument	M220	ME220	S220	E220 Evolution	E220	LE220
Target Size (bp)	200-700	200-700	200-700	200-700	200-700	200-700
PIP	75	75	105	105	105	300
Duty Factor (%)	5	5	2	2	2	15
CPB	200	1000	200	200	200	200
Treatment Time (minutes)	2-12	2-12	2-12	2-12	2-12	15-30
Setpoint Temperature (°C) <sub>1</sub>	7	7	6	6	6	6
Min/Max Temperature (°C)	4/10	6/12	3/9	3/9	3/9	3/9
Sample Input	<100,000	<100,000	<100,000	<100,000	<100,000	<100,000
AFA Intensifier Required <sub>2</sub>	NA	NA	Integrated	Yes	Yes	NA
Water Level (run) <sub>3</sub>	Full	9	12	6	6	6
Sample Volume (µl) <sub>4</sub>	130	130	130	130	130	130
<b>Important Notes</b>						
<ol style="list-style-type: none"> <li>1. If using the S220, E220 Evolution, E220, or LE220, set the temperature on the external chiller 3°C below the setpoint temperature for the run. The min/max is set in SonoLab</li> <li>2. If intensifier is required, please ensure PN 500141 is used</li> <li>3. Water level should always be 1mm below the neck of the microTUBE-130 cap</li> <li>4. Always fill the microTUBE-130 with 130 µl of sample</li> </ol>						

## Appendix B: Chromatin Shearing Efficiency Analysis Protocol

1. Take a 130  $\mu\text{L}$  aliquot of the sheared sample and transfer to 0.6 mL microcentrifuge tube.
2. Add 5  $\mu\text{L}$  of RNase A (10 mg/mL) and incubate at 37 °C for 30 min.
3. Add 5  $\mu\text{L}$  of Proteinase K (10 mg/mL) and reverse crosslink by heating at 65 °C overnight in a PCR cycler with a heated lid.
4. Purify DNA using either a commercial column based kit (*e.g.*, Qiagen QIAquick PCR Purification Kit, Cat. No. 28104), or phenol-chloroform extraction and ethanol precipitation.
5. Elute from column, or resuspend pellet with 25  $\mu\text{L}$  of elution buffer (10 mM Tris-HCl, pH 8.5).
6. 1 $\mu\text{L}$  of purified DNA can be analyzed on an Agilent 2100 BioAnalyzer 12K chip to provide a more accurate representation of the shearing size range and distribution.
7. Alternatively, an aliquot of the sample can be run on a High Sensitivity Agilent 2100 chip.

## Appendix C: Additional Notes

1. The treatment settings listed in this document are recommended guidelines. Actual results may vary depending on the cell type and mass.
2. The Covaris process uses high intensity focused ultrasonic (HIFU) energy and as such is influenced by objects in the acoustic path from the transducer surface to the fluid sample. For example, particles and bubbles in the water bath may scatter the acoustic energy from the sample. Replace the bath water on a daily basis and ensure that appropriate time has been allowed for degassing and water bath temperature to stabilize prior to use of the instrument.
3. Bubbles in the sample fluid in the tube may diminish the acoustic dose effectiveness. Be sure to fill the tubes slowly with the recommended volumes and avoid the use of additional detergents that may induce foaming.

## REFERENCES

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